

# Korup National Park

CMNTIPA035



Country: **Cameroon**  
Administrative region: **Southwest (Region)**  
Central co-ordinates: **5.20000 N, 8.90000 E**  
Area: **1873km<sup>2</sup>**

## Qualifying IPA criteria

A(i)

## IPA assessment rationale

Korup National Park qualifies as a potential IPA under criterion A(i) due to the presence of a very high number of globally threatened species, several of which are endemic or nearly so. It also likely qualifies under criterion B(ii) for its many range restricted species, B(iii) for its richness in wild harvested species, and criterion C(iii) as a representative of its habitat type.

## Site description

Korup National Park, in Southwest Region (Manyu and Ndiang divisions), Cameroon, was established in 1986 following research by primatologists and NGOs such as WWF from the late 1970s. It expands on the former Korup Forest Reserve, which dates to the

1930s. The National Park's borders encompass 1,259 km<sup>2</sup> of lowland forest rainforest, with another 600 km<sup>2</sup> demarcated through a 3 km buffer zone all around the perimeter except where it joins the border with Nigeria. A 100 km<sup>2</sup> core zone in the south near Mundemba is further demarcated, and agencies and government have also targeted for development and conservation work a much larger Korup Project Area, incorporating Ejagham Forest Reserve to the north, Nta Ali reserve in the northeast, Rumpi Hills in the east, and land in the south up to the mangrove forest of Ndongere proposed national park. The boundaries of the IPA proposed here are constituted by the Korup National Park and buffer zone. The park has its official entrance, the iconic suspension bridge spanning the river Ndiang, in the south, close to the town of Mundemba. The towns of Nguti and Manyemen serve the northeast part. Principal access is along a 120 km long unpaved road from Kumba to Mundemba, and can be difficult in the wet season.

## Botanical significance

Korup is situated within the Biafran rain forest block (Letouzey, 1968), a part of the Guineo-Congolian Regional Centre of Endemism (White, 1983). The floral and faunal diversity of the Biafran rain forest zone is very high and there are many endemics (Gartlan, 1994). Southwestern Cameroon has been indicated to have the highest diversity of both species and genera per degree square in tropical Africa (Barthlott et al., 1996; Dagallier et al., 2020). The

National Park is nearly completely covered in lowland and sub-montane evergreen rain forest and is one of the most important botanical areas in tropical Africa (Gartlan, 1994). The site is relatively well studied compared to other areas of coastal forest in neighbouring Nigeria and Cameroon, and is known to harbour very high plant diversity with around 3,500 vascular plant species (Gartlan, 1994). Many species are endemic to the site itself or known only from Korup and a few other nearby sites such as Mt Cameroon, Rumpi Hills, Ejagham Forest Reserve or Oban Hills in Nigeria. Exceptionally high rainfall and particular edaphic properties may partly explain the botanical richness. Korup is also considered a forest refugium where the climate has been relatively stable during the Pleistocene, enabling the forest to persist during periods with a generally cooler and drier climate (Maley, 1998; Sosef, 1994). The tree species composition in the southern part of the park shows a remarkably low presence of pioneer species, indication that this part of the park has been little affected by natural and anthropogenic disturbances (X.M. van der Burgt, unpublished data). Elsewhere in the park, some areas with secondary forest occur (Gartlan, 1994).

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## Habitat and geology

Korup has a two-season climate, with the wet season running between March and November. It is one of the wettest areas in tropical Africa. In the south of the park, total annual rainfall averaged 5,382 mm over a 14 year period, with the months December to February averaging c.100 mm per month, and July and August each averaging c. 1,000 mm per month. Rainfall is considerably less in the north of the park, averaging 2,200 mm, although measurements were from a different source and averaged over a much shorter period (Rodewald et al., 1994). Mean monthly temperatures range from c. 24-30° C and mean peak temperatures range seasonally between c. 30-32° C, with a diurnal range of 10° C (Chuyong, 2004; Thomas et al., 2015).

The area is underlain by ancient metamorphic basement rocks, with some granitic outcrops and boulders (Letouzey 1985; Burgt, 2018). Soils are classified as predominantly yellow ferralitic soils over acid rocks according to Vallerie (1970) or nitisols according to Yerima & Van Ranst (2005). They are typically described as strongly weathered, sandy, acidic and infertile, with deficient phosphate and potassium (Burgt, 2018; Chuyong, 2004; Newbery et al., 2004). Periodically inundated soils occur along streams and rivers (Burgt, 2018).

Three major rivers flow through the park, taking different routes to meet the sea in the same approximate area. The Ndian river flows south from Ejagham, through the middle of the park, then forms the eastern boundary and is joined by tributaries from the Rumpi Hills area before continuing to meet the sea at the Rio del Rey. The Korup river forms the western boundary of the park and part of the international border with Nigeria, flowing south to join the Cross River estuary. The Munaya river flows north from the Rumpi Hills through northwest Korup, to join the Cross River which also eventually turns southwards towards the Bight of Bonny. Nearly 50% of the national park is below 360 m altitude (Rodewald et

al., 1994). Mt Rana, in the middle of the park, is the highest point at 1,080 m, and there is also steep terrain in the northwest part. Closed-canopy, lowland, coastal evergreen rainforest covers almost the entire site but patches of periodically inundated rainforest occur along watercourses and some sub-montane forest appear on the highest points; tree species from the Leguminosae-Detarioideae are locally abundant and sometimes dominant (Newbery et al., 2004). There are also a few granite inselbergs and steep granite slopes with shrubby vegetation and bare rock. Some areas have been cleared for cocoa farming, and extensive areas in the buffer zone for palm oil (Diaw et al., 2003; Siewe et al., 2017).

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## Conservation issues

Korup National Park represents one of the first attempts in West or Central Africa at integrated conservation and development (Malleison, 2002). Agencies including WWF-UK, WCS, UK ODA (now FCDO), the German development agency GTZ (now GIZ) and the European Commission were involved with the Cameroon government in the establishment and funding of the national park and associated Korup Project (Oates et al., 2004). Implementation has been criticised for the treatment of villagers inside the park (Diaw et al., 2003; Siewe et al., 2017). Five villages remain within the park boundary (Erat, Bera, Esukutan, Ikenge, Bareka Batanga) and another 25 are situated within the buffer zone where development projects have been focused (Siewe et al., 2017; Malleison, 2002). One village, Ikondo-Kondo has been relocated, in 2000, to a new location, while other efforts have been abandoned (Malleison, 2002). The relocation policy has been controversial and villagers have reportedly been left feeling angry and betrayed (Malleison, 2002; Diaw et al., 2003). Since 2017, a new collaborative management approach has been attempted and villagers and other stakeholders have provided input to the 2017-2021 management plan (Shu & Ebuja, 2018). This development has also led to permanent use zones (PUZs), legalising the status of two of the remaining 5 villages within the park. As of 2017 PUZs had yet to be implemented for the remaining villages (Shu & Ebuja, 2018).

Up to 8% of the national park area has apparently been cultivated by villagers, with cultivation increasing in response to abandonment of relocation plans and restrictions on use of the forest for hunting and other resources (Malleison, 2002). Cocoa farming is the main agricultural activity and almost universally practised by villagers (Siewe et al., 2017). However, villagers continue to utilise the forest, and bushmeat hunting was reported as continuing, with sales across the Nigerian border evading policing efforts (Malleison, 2002; Siewe et al., 2017). Large scale oil palm cultivation is evident in the buffer zone, logging is reported to remain a major threat nearby, and numerous forestry concessions surround the park and overlap the buffer zone (KBA partnership, 2020; WRI, 2020). The Anglophone crisis has interrupted conservation work and brought significant numbers of people seeking refuge within the forest, likely increasing agricultural and hunting activity (Linder et al., 2019).

Korup National Park is adjacent to the protected Cross River National Park in Nigeria, which enhances the size and importance of

the overall habitat. There are also connections with other IPAs and conservation sites in Cameroon, including Ejagham and Nta Ali Forest Reserves, Rumpi Hills and Banyang Mbo Wildlife Reserve, Ndongere proposed National Park and the Mt Cameroon area. Connecting corridors are, however, increasingly threatened by development which may limit dispersal and migration. Populations of mammals have declined significantly as a result of poaching, and this will negatively affect the recruitment of many plant species in the forest that are reliant on mammal dispersers (Abernethy et al., 2013; Estrada et al., 2017).

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### **Site assessor(s)**

Bruce Murphy, Royal Botanic Gardens, Kew

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## IPA criterion A species

SPECIES	QUALIFYING SUB-CRITERION	≥ 1% OF GLOBAL POPULATION	≥ 5% OF NATIONAL POPULATION	1 OF 5 BEST SITES NATIONALLY	ENTIRE GLOBAL POPULATION	SOCIO-ECONOMICALLY IMPORTANT	ABUNDANCE AT SITE
<i>Aframomum tchoutoui</i> D.J.Harris & Wortley	A(i)	✓	✓	✓	–	–	
<i>Afrothimia hydra</i> Sainge & T.Franke	A(i)	✓	✓	✓	–	–	
<i>Ancistrocladus korupensis</i> D.W.Thomas & Gereau	A(i)	✓	✓	✓	–	–	
<i>Aporrhiza multijuga</i> Gilg	A(i)	✓	✓	✓	–	–	
<i>Begonia quadrialata</i> Warb. subsp. <i>dusenii</i> (Warb.) Sosef	A(i)	✓	✓	✓	–	–	
<i>Beilschmiedia jacques-felixii</i> Robyns & R.Wilczek	A(i)	✓	✓	✓	–	–	
<i>Cassipourea korupensis</i> Kenfack & Sainge	A(i)	✓	✓	✓	✓	–	
<i>Cola mamboana</i> Kenfack & Sainge	A(i)	✓	✓	✓	–	✓	
<i>Cola suboppositifolia</i> Cheek	A(i)	✓	✓	✓	–	✓	
<i>Cola zemagoana</i> Kenfack & D.W.Thomas	A(i)	✓	✓	✓	✓	✓	
<i>Cryptosepalum korupense</i> Burgt	A(i)	✓	✓	✓	–	–	
<i>Deinbollia angustifolia</i> D.W.Thomas	A(i)	✓	✓	✓	✓	–	
<i>Deinbollia saligna</i> Keay	A(i)	✓	✓	✓	–	–	
<i>Deinbollia unijuga</i> D.W.Thomas	A(i)	✓	✓	✓	–	–	
<i>Dichapetalum korupinum</i> Breteler	A(i)	✓	✓	✓	✓	–	
<i>Dichapetalum letouzeyi</i> Breteler	A(i)	✓	✓	✓	✓	–	

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<i>Didelotia korupensis</i> Burgt	A(i)	✓	✓	✓	✓	–	
<i>Empogona talbotii</i> (Wernham) Tosh & Robbr.	A(i)	✓	✓	✓	–	–	
<i>Gardenia epiphytica</i> Jongkind	A(i)	✓	✓	✓	–	–	
<i>Gluema korupensis</i> Burgt	A(i)	✓	✓	✓	–	–	
<i>Guaduella humilis</i> Clayton	A(i)	✓	✓	✓	–	–	
<i>Hugonia batesii</i> De Wild.	A(i)	✓	✓	✓	–	–	
<i>Hymenostegia bakeriana</i> Hutch. & Dalziel	A(i)	✓	✓	✓	–	–	
<i>Ixora delicatula</i> Keay	A(i)	✓	✓	✓	–	–	
<i>Justicia tenuipes</i> S.Moore	A(i)	✓	✓	✓	–	–	
<i>Korupodendron songweanum</i> Litt & Cheek	A(i)	✓	✓	✓	–	–	
<i>Lecomtedoxa plumosa</i> Burgt	A(i)	✓	✓	✓	✓	–	
<i>Memecylon korupense</i> R.D.Stone	A(i), A(iii), A(iv)	✓	✓	✓	✓	–	
<i>Memecylon rheophyticum</i> R.D.Stone, Ghogue & Cheek	A(i), A(iii), A(iv)	✓	✓	✓	✓	–	
<i>Pavetta baconiella</i> Bremek.	A(i)	✓	✓	✓	✓	–	
<i>Pavetta grossissima</i> S.D.Manning	A(i)	✓	✓	✓	✓	–	
<i>Physacanthus talbotii</i> S.Moore	A(i)	✓	✓	✓	–	–	
<i>Piptostigma oyemense</i> Pellegr.	A(i)	✓	✓	✓	–	–	
<i>Placodiscus caudatus</i> Pierre ex Pellegr.	A(i)	✓	✓	✓	–	–	

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<i>Psychotria elephantina</i> Lachenaud & Cheek	A(i)	✓	✓	✓	–	–	
<i>Rinorea thomasii</i> Achound.	A(i)	✓	✓	✓	–	–	
<i>Salacia lenticellosa</i> Loes. ex Harms	A(i)	✓	✓	✓	–	–	
<i>Staurogyne kamerunensis</i> (Engl.) Benoist subsp. <i>calabarensis</i> Champl.	A(i)	✓	✓	✓	–	–	
<i>Talbotiella korupensis</i> Mackinder & Wieringa	A(i)	✓	✓	✓	–	–	
<i>Tessmannia korupensis</i> Burt	A(i)	✓	✓	✓	–	–	
<i>Tricalysia achoundongiana</i> Robbr., Sonké & Kenfack	A(i)	✓	✓	✓	–	–	
<i>Vepris heterophylla</i> (Engl.) Letouzey	A(i)	–	✓	✓	–	–	
<i>Vepris letouzeyi</i> Onana	A(ii)	✓	✓	✓	–	–	
<i>Boutiquea platypetala</i> (Engl. & Diels) Le Thomas	A(i)	✓	✓	✓	–	–	
<i>Isolona pleurocarpa</i> Diels	A(i)	✓	✓	✓	–	–	
<i>Landolphia maxima</i> (K.Schum. ex Hallier f.) Pichon	A(i)	✓	✓	✓	–	–	
<i>Hoplostigma pierreanum</i> Gilg	A(i), A(iii)	✓	✓	✓	–	–	
<i>Salacia lucida</i> Oliv.	A(i)	✓	✓	✓	–	–	
<i>Grossera major</i> Pax	A(i)	✓	–	✓	–	–	
<i>Eurypetalum unijugum</i> Harms	A(i)	✓	✓	✓	–	–	

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<i>Strychnos gnetifolia</i> Gilg ex Onochie & Hepper	A(i)	✓	✓	✓	–	–	
<i>Phyllanthus dusenii</i> Hutch.	A(i)	✓	✓	✓	–	–	
<i>Drypetes preussii</i> (Pax) Hutch.	A(i)	–	–	✓	–	–	
<i>Drypetes staudtii</i> (Pax) Hutch.	A(i)	✓	–	✓	–	–	
<i>Belonophora ongensis</i> S.E.Dawson & Cheek	A(i), A(iii)	✓	✓	✓	–	–	
<i>Belonophora talbotii</i> (Wernham) Keay	A(i), A(iii)	✓	✓	✓	–	–	
<i>Psychotria densinervia</i> (K.Krause) Verdc.	A(i), A(iii)	–	–	✓	–	–	
<i>Psychotria lanceifolia</i> K.Schum.	A(i)	✓	✓	✓	–	–	
<i>Tricalysia ferorum</i> Robbr.	A(i), A(iii)	✓	✓	✓	–	–	
<i>Deinbollia pycnophylla</i> Gilg ex Engl.	A(i)	✓	✓	✓	–	–	
<i>Placodiscus angustifolius</i> Radlk.	A(i)	✓	✓	✓	–	–	
<i>Dicranolepis polygaloides</i> Gilg ex H.Pearson	A(i)	✓	–	✓	–	–	
<i>Afrofittonia silvestris</i> Lindau	A(i)	✓	✓	✓	–	–	
<i>Asystasia lindauiana</i> Hutch. & Dalziel	A(i)	✓	✓	✓	–	–	
<i>Schefflera mannii</i> (Hook.f.) Harms	A(i)	✓	–	✓	–	–	
<i>Berlinia korupensis</i> Mackinder & Burgt	A(i)	✓	✓	✓	–	–	
<i>Gilbertiodendron newberyi</i> Burgt	A(i)	✓	✓	✓	–	✓	
<i>Microberlinia bisulcata</i> A.Chev.	A(i)	✓	✓	✓	–	✓	

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<i>Chazaliella obanensis</i> (Wernham) Petit & Verdc.	A(i)	✓	–	–	–	–	
<i>Hymenostegia talbotii</i> Baker f.	A(i)	✓	✓	✓	–	✓	
<i>Hymenostegia viridiflora</i> Mackinder & Wieringa	A(i)	✓	✓	✓	–	–	
<i>Costus albiflos</i> Maas & H.Maas	A(i)	✓	✓	✓	–	–	
<i>Vitex lokundjensis</i> W.Piep.	A(i)	✓	✓	✓	–	–	
<i>Anthonotha xanderi</i> Breteler	A(i)	✓	✓	✓	–	–	
<i>Begonia oxyanthera</i> Warb.	A(i)	✓	–	–	–	✓	
<i>Memecylon dasyanthum</i> Gilg & Ledermann ex Engl.	A(i)	✓	✓	✓	–	✓	
<i>Corymborkis minima</i> P.J.Cribb	A(i)	✓	✓	✓	–	–	
<i>Afropectinariella pungens</i> (Schltr.) M.Simo & Stévant	A(i)	✓	✓	✓	–	–	
<i>Anthocleista scandens</i> Hook.f.	A(i)	✓	–	–	–	–	
<i>Leonardoxa africana</i> (Baill.) Aubrév. subsp. <i>letouzeyi</i> McKey	A(i)	✓	✓	✓	–	–	
<i>Begonia preussii</i> Warb.	A(i)	✓	–	✓	–	–	
<i>Napoleonaea egertonii</i> Baker f.	A(i)	✓	–	✓	–	–	
<i>Salacia lehmbachii</i> Loes var. <i>pes-ranulae</i> N.Hallé	A(i)	✓	–	✓	–	–	
<i>Jollydora glandulosa</i> G.Schellenb.	A(i)	✓	✓	✓	–	–	
<i>Calochone acuminata</i> Keay	A(i)	✓	✓	✓	–	✓	
<i>Angylocalyx talbotii</i> Baker f. ex	A(i)	–	–	✓	–	–	



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<i>Hutch. &amp; Dalziel</i>							
<i>Antrocaryon micrastrer</i> A.Chev. & Guillaumin	A(i)	–	–	✓	–	✓	
<i>Cola megalophylla</i> Brenan & Keay	A(i)	✓	✓	✓	–	✓	
<i>Crateranthus talbotii</i> Baker f.	A(i)	–	–	✓	–	–	
<i>Garcinia staudtii</i> Engl.	A(i)	✓	–	✓	–	–	
<i>Rhodognaphalon brevicuspe</i> (Sprague) Roberty	A(i)	–	–	✓	–	✓	
<i>Warneckea austro-occidentalis</i> R.D.Stone	A(i)	✓	✓	✓	–	–	
<i>Ancistrocladus grandiflorus</i> Cheek	A(i)	✓	✓	✓	–	–	
<i>Cola nigerica</i> Brenan & Keay	A(i)	✓	✓	✓	–	–	
<i>Diospyros korupensis</i> Gosline	A(i)	✓	✓	✓	–	✓	
<i>Amanoa strobilacea</i> Müll.Arg.	A(i)	–	✓	✓	–	–	
<i>Cola praecuta</i> Brenan & Keay	A(i)	✓	✓	✓	–	–	
<i>Liparis gracilentis</i> Dandy	A(i)	✓	✓	✓	–	–	
<i>Sabicea medusula</i> K.Schum. ex Wernham	A(i)	✓	✓	✓	–	–	
<i>Uvariopsis korupensis</i> Gereau & Kenfack	A(i)	✓	✓	✓	–	–	
<i>Aframomum plicatum</i> D.J.Harris & Wortley	A(i)	✓	✓	✓	–	–	
<i>Begonia prismatocarpa</i> Hook. subsp. <i>delobata</i> Sosef	A(i)	✓	✓	✓	–	–	
<i>Cuviera talbotii</i> (Wernham) Verdc.	A(i)	✓	–	✓	–	–	

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<i>Macropodiella pellucida</i> (Engl.) C.Cusset	A(i)	✓	✓	✓	–	–	
<i>Placodiscus opacus</i> Radlk.	A(i)	✓	–	✓	–	–	
<i>Psychotria minimicalyx</i> K.Schum	A(i)	✓	–	✓	–	–	
<i>Trichostachys interrupta</i> K.Schum.	A(i)	✓	✓	✓	–	–	
<i>Vepris trifoliolata</i> (Engl.) Mziray	A(i)	✓	✓	✓	–	–	
<i>Tricalysia lejolyana</i> Sonké & Cheek	A(i)	✓	✓	✓	–	✓	
<i>Allexis obanensis</i> Violaceae	A(i)	✓	✓	✓	–	–	
<i>Memecylon candidum</i> , Melastomataceae	A(i)	–	–	✓	–	–	
<i>Gaertnera letouzeyi</i> Malcomber	A(i)	✓	✓	✓	–	–	
<i>Allexis cauliflora</i>	A(i)	✓	✓	✓	–	–	
<i>Amanoa bracteosa</i> Planch.	A(i)	✓	✓	✓	–	–	
<i>Baillonella toxisperma</i> Pierre	A(i)	–	✓	✓	–	✓	
<i>Daniellia oblonga</i> Oliv.	A(i)	✓	✓	✓	–	–	
<i>Diospyros crassiflora</i> Hiern	A(i)	–	–	✓	–	✓	
<i>Garcinia kola</i> Heckel	A(i)	–	–	✓	–	✓	
<i>Lepalea thompsonii</i> (Sprague & Hutch.) E.J.M.Koenen & J.J.de Wilde	A(i)	–	–	✓	–	✓	
<i>Lophira alata</i> Banks ex Gaertn.f.	A(i)	–	–	✓	–	✓	
<i>Magnistipula cuneatifolia</i> Hauman	A(i)	✓	✓	✓	–	–	

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<i>Magnistipula multinervia</i>	A(i), A(iii)	✓	✓	✓	✓	–	
<i>Manilkara lososiana</i>	A(i)	✓	✓	✓	✓	–	
<i>Turraeanthus mannii</i> Baill.	A(i)	–	–	–	–	✓	
<i>Berlinia hollandii</i> Hutch. & Dalziel	A(i)	✓	✓	✓	–	–	
<i>Millettia laurentii</i> de Wild.	A(i)	–	✓	✓	–	–	
<i>Salacia nigra</i> Cheek	A(i)	✓	✓	✓	–	–	
<i>Salacia volubilis</i> Loes. & H.J.P.Winkl.	A(i)	✓	–	–	–	–	
<i>Magnistipula butayei</i> De Wild. subsp. <i>korupensis</i> Burgt	A(iii)	✓	✓	✓	✓	–	
<i>Afrothismia korupensis</i> Sainge & T.Franke	A(i)	✓	✓	✓	✓	–	
<i>Bulbophyllum subligaculiferum</i> J.J.Verm.	A(i)	–	–	–	–	–	
<i>Aframomum makandensis</i> Dhetchuvi	A(i)	✓	✓	✓	–	–	
<i>Anisophyllea neopurpurascens</i> Li Bing Zhang, Xin Chen & H.He	A(i)	✓	✓	✓	–	–	
<i>Craterispermum capitatum</i> Taedoumg & De Block	A(i)	✓	✓	✓	–	–	
<i>Garcinia afzelii</i> Engl.	A(i)	–	–	✓	–	–	
<i>Neolemonniera ogouensis</i> (Dubard) Heine	A(i)	✓	✓	✓	–	–	
<i>Cola brevipes</i> Malvaceae	A(i)	✓	✓	✓	–	–	
<i>Globulostylis minor</i> Wernham	A(iii)	✓	✓	✓	–	–	
<i>Piptostigma longepilosum</i>	A(i)	✓	✓	✓	–	–	

SPECIES	QUALIFYING SUB-CRITERION	≥ 1% OF GLOBAL POPULATION	≥ 5% OF NATIONAL POPULATION	1 OF 5 BEST SITES NATIONALLY	ENTIRE GLOBAL POPULATION	SOCIO-ECONOMICALLY IMPORTANT	ABUNDANCE AT SITE
<i>Piptostigma macrophyllum</i> Ghogue, Sonké & Couvreur	A(i)	✓	✓	✓	–	–	
<i>Psychotria camerunensis</i> E.M.A.Petit	A(i)	✓	–	–	–	–	
<i>Pterygota bequaertii</i> De Wild.	A(i)	–	–	–	–	–	
<i>Terminalia ivorensis</i> A.Chev.	A(i)	–	✓	✓	–	–	
<i>Begonia rubromarginata</i> Gilg	A(i)	✓	✓	✓	–	–	
<i>Allophylus nigericus</i> Baker f.	A(iv)	✓	✓	✓	–	–	
<i>Callichilia basileis</i> Beentje	A(iv)	✓	✓	✓	–	–	
<i>Cassipourea atanganae</i> Kenfack	A(iii)	✓	✓	✓	✓	–	
<i>Chytranthus micranthus</i> Gilg ex Radlk.	A(iv)	✓	✓	✓	–	–	
<i>Costus acutissimus</i> Maas & H.Maas	A(iii)	✓	✓	✓	–	–	
<i>Crotonogynopsis korupensis</i> Kenfack & D.W.Thomas	A(iv)	✓	✓	✓	–	–	
<i>Cuviera trilocularis</i> Hiern	A(iv)	✓	✓	✓	–	–	
<i>Psychotria korupensis</i> O.Lachenaud	A(iii)	✓	✓	✓	✓	–	
<i>Scaphopetalum parvifolium</i> Baker f.	A(i)	✓	✓	✓	–	–	
<i>Scyphosyce pandurata</i> Hutch.	A(iv)	✓	✓	✓	–	–	

## IPA criterion C qualifying habitats

HABITAT	QUALIFYING SUB-CRITERION	≥ 5% OF NATIONAL RESOURCE	≥ 10% OF NATIONAL RESOURCE	1 OF 5 BEST SITES NATIONALLY	AREAL COVERAGE AT SITE
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## General site habitats

GENERAL SITE HABITAT	PERCENT COVERAGE	IMPORTANCE
Forest - Subtropical/Tropical Moist Lowland Forest	90	Major
Forest - Subtropical/Tropical Swamp Forest	1	Minor
Forest - Subtropical/Tropical Moist Montane Forest	3	Minor

## Land use types

LAND USE TYPE	PERCENT COVERAGE	IMPORTANCE
Nature conservation	90	Major
Agriculture (arable)	10	Minor

## Threats

THREAT	SEVERITY	TIMING
Agriculture & aquaculture - Annual & perennial non-timber crops - Shifting agriculture	Medium	Ongoing - increasing
Agriculture & aquaculture - Annual & perennial non-timber crops - Small-holder farming	Medium	Ongoing - increasing
Agriculture & aquaculture - Annual & perennial non-timber crops - Agro-industry farming	Medium	Ongoing - increasing
Biological resource use - Hunting & collecting terrestrial animals	High	Ongoing - trend unknown
Biological resource use - Logging & wood harvesting	Medium	Ongoing - trend unknown

## Protected areas

PROTECTED AREA NAME	PROTECTED AREA TYPE	RELATIONSHIP WITH IPA	AREAL OVERLAP
Korup National Park	National Park	protected/conservation area matches IPA	1873

## Conservation designation

DESIGNATION NAME	PROTECTED AREA	RELATIONSHIP WITH IPA	AREAL OVERLAP
Korup National Park	Important Bird Area	IPA encompasses protected/conservation area	1259
Korup National Park	Key Biodiversity Area	IPA encompasses protected/conservation area	1259

## Management type

MANAGEMENT TYPE	DESCRIPTION	YEAR STARTED	YEAR FINISHED
Site management plan in place	Under the new collaborative management approach implemented by MINEF through the Programme for Sustainable Management of Natural Resources in the South West Region (PSMNR-SWR), the 32 villages and other stakeholders were reportedly involved in creation of the latest management plan.	2017	2021

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